Fresh as an Exitron: A Flower-specific Splice Variant of AUXIN RESPONSE FACTOR8 Helps Shape the Stamen

Eukaryotic genes contain protein-coding exons interspersed with non-coding introns. While introns are usually spliced out of mRNA (often in conjunction with various exons), intron retention usually causes mRNA to remain in the nucleus instead of being exported to the cytoplasm for translation. This process stalls gene expression at a particular stage, tissue, or condition, thereby regulating development or stress responses. To complicate matters, some exons contain exitrons (Marguez et al., 2015). These intronlike regions allow intraexonic protein-coding sequences to be differentially spliced, yielding different mRNAs. However, in contrast to canonical intron retention, transcripts with retained exitrons exit the nucleus for translation into different proteins in a tissuespecific manner.

Such plasticity no doubt comes in handy during flower development, when relatively few transcription factors orchestrate the expression of scores of genes. Thousands of transcripts generated by alternative splicing, including intron retention, are differentially expressed during *Arabidopsis thaliana* flower development (Wang et al., 2014).

For example, the splice variant *ARF8.2* has been found for *AUXIN RESPONSE FACTOR8*, encoding a transcription factor that binds to auxin response elements in the promoters of target genes to control their expression. Auxin in developing stamens upregulates *AUX/IAA19* levels and controls the degradation of this transcriptional repressor, thus preventing ARF8 from binding to its targets. This repression results in filament elongation, explaining the short stamens of the *arf8* mutant.

While exploring possible roles for *ARF8.2* in stamen development, **Ghelli et al.** (2018) came across an intriguing new splice variant, *ARF8.4*. *ARF8.4* is exclusively expressed in stamens during late development and is translated and efficiently transported to the nucleus, as revealed by transient expression analysis, indicating that it contains a retained exitron. To investigate its role, the authors produced Arabidopsis *arf8* lines harboring the full-length *ARF8.1*,

ARF8.2, and ARF8.4 coding sequences driven by an inducible promoter (ARF8.1oxarf8, ARF8.2ox-arf8, and ARF8.4ox-arf8, respectively). Only ARF8.4 fully complemented the short-stamen phenotype of arf8, pointing to its role in stamen elongation. AUX/IAA19 was upregulated in ARF8.4oxarf8 plants, perhaps explaining their increased stamen length, as confirmed by chromatin

8.2ox8.4ox





Co-expression of *ARF8.4* and *ARF8.2* **Causes Precocious Anther Dehiscence.** Flowers from mock (top) and inducer (bottom)-treated plants co-expressing *ARF8.2* and *ARF8.4* (8.20x8.40x). (*Reprinted from Napoli et al.* [2018], *Figure 5A.*)

immunoprecipitation-qPCR.

Wild-type and arf8 plants coexpressing ARF8.2 and ARF8.4 showed precocious anther dehiscence (see figure), whereas ARF8.2ox and ARF8.4ox plants did not. Anther dehiscence is linked to the indirect activation of the jasmonic acid (JA) biosynthesis gene DEFECTIVE IN ANTHER DEHISCENCE1 (DAD1) by ARF8. ARF8.2ox flowers had increased DAD1 transcript levels, whereas ARF8.4ox flowers did not, suggesting that ARF8.2 regulates anther dehiscence by affecting JA biosynthesis. Histological analysis suggested that ARF8.4 regulates lignin deposition, an early step in anther dehiscence. This process is controlled by the transcription factor MYB26, whose expression is negatively regulated by auxin (Cecchetti et al., 2013). Indeed, ARF8.4 binds strongly to the MYB26 promoter and regulates its expression, whereas ARF8.2 has only minor effects on this gene.

Therefore, two *ARF8* splice variants control different steps in stamen development, with the evolutionarily conserved exitron-retaining splice variant *ARF8.4* playing crucial roles in this process. Whether this fresh take on flower development represents a general mechanism in plant organ development remains to be explored.

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